

Environmental Contamination with Cyclophosphamide, Ifosfamide, and Methotrexate: A Study of 51 Canadian Centres

Alexia Janes, Cynthia Tanguay, Nicolas J Caron, and Jean-François Bussi eres

ABSTRACT

Background: Occupational exposure to hazardous drugs may lead to adverse reproductive effects. There is no safe exposure limit for health care professionals.

Objectives: To monitor levels of cyclophosphamide, ifosfamide, and methotrexate contamination in oncology pharmacy and patient care areas in Canadian health care institutions.

Methods: The study was conducted in 2014. Hospitals with at least 50 acute care beds were invited to participate. At each participating centre, 12 standardized sites (6 in pharmacy areas and 6 in patient care areas) were sampled. The samples were analyzed for the presence of cyclophosphamide, ifosfamide, and methotrexate by ultra-performance liquid chromatography tandem mass spectrometry technology. The limits of detection were 0.36 pg/cm² for cyclophosphamide, 0.95 pg/cm² for ifosfamide, and 0.97 pg/cm² for methotrexate. Descriptive statistical analyses were performed to determine the median, 75th percentile, and maximum levels.

Results: Fifty-one hospitals participated in this descriptive study, and a total of 584 samples were quantified. Overall, 294 (50%) of the samples were positive for cyclophosphamide, 125 (21%) for ifosfamide, and 54 (9%) for methotrexate. The most frequently contaminated sampling sites in pharmacy areas were the front grille inside the hood and the floor in front of the hood and, in patient care areas, the armrest and outpatient clinic counter. The 75th percentiles for surface concentration were 10.8 pg/cm² for cyclophosphamide, 1.59 pg/cm² for ifosfamide, and below the limit of detection for methotrexate.

Conclusions: Relative to 3 other multicentre studies conducted in Quebec over the past few years, the proportion of positive samples remained constant. Nonetheless, the 75th percentile surface concentration of antineoplastic drugs has been decreasing and seems to have reached a plateau. Local (country-specific or region-specific) and attainable goals for surface contamination with hazardous drugs should be set annually, so long as no health-based limit is known.

Keywords: occupational exposure, environmental monitoring, antineoplastic agents, cyclophosphamide, ifosfamide, methotrexate

R SUM 

Contexte : L'exposition professionnelle   des m dicaments dangereux peut causer des effets ind sirables sur la reproduction. Aucune limite d'exposition s curitaire n'est  tablie pour les professionnels de la sant .

Objectifs :  valuer les taux de cyclophosphamide, d'ifosfamide et de m thotrexate dans la pharmacie d'oncologie et dans les unit s de soins des  tablissements de sant  canadiens.

M thodes : L' tude s'est d roul e en 2014. Les h pitaux disposant d'au moins 50 lits de soins de courte dur e ont  t  invit s   participer. Dans chacun des  tablissements participants, des  chantillons ont  t  pr lev s dans 12 zones pr d termin es : 6 dans les pharmacies et 6 dans les unit s de soins. On a ensuite analys  les  chantillons par chromatographie liquide   tr s haute performance coupl e   la spectrom trie de masse en tandem afin de d tecter la pr sence de cyclophosphamide, d'ifosfamide et de m thotrexate. Le seuil de d tection  tait de 0,36 pg/cm² pour la cyclophosphamide, de 0,95 pg/cm² pour l'ifosfamide et de 0,97 pg/cm² pour le m thotrexate. Des analyses statistiques descriptives ont  t  effectu es afin de d terminer la m diane, le 75^e percentile et les taux maximums.

R sultats : Au total, 51 h pitaux ont particip    cette  tude descriptive et 584  chantillons ont  t  quantifi s. Dans l'ensemble, 294 (50 %)  chantillons  taient positifs pour la cyclophosphamide, 125 (21 %) pour l'ifosfamide et 54 (9 %) pour le m thotrexate. Les zones les plus fr quemment contamin es  taient : en pharmacie, la grille avant dans la hotte et le sol devant la hotte; dans les unit s de soins, les accoudoirs et le comptoir des cliniques de consultation externe. Le 75^e percentile de la concentration de surface  tait de 10,8 pg/cm² pour la cyclophosphamide, 1,59 pg/cm² pour l'ifosfamide et sous le seuil de d tection pour le m thotrexate.

Conclusions : Comparativement   trois autres  tudes multicentriques men es au Qu bec au cours des derni res ann es, la proportion de pr l vements positifs demeure la m me. Toutefois, le 75^e percentile de la concentration de surface d'antineoplasiques a diminu  et semble avoir atteint un plateau. Des objectifs locaux (pour le pays ou selon les r gions) et r alisables de contamination de surface par des m dicaments dangereux devraient  tre  tablis chaque ann e, et ce, tant qu'aucune limite fond e sur les crit res li s   la sant  ne sera pas d termin e.

Mots cl s : exposition professionnelle, surveillance environnementale, antineoplasiques, cyclophosphamide, ifosfamide, m thotrexate

Can J Hosp Pharm. 2015;68(4):279-89

INTRODUCTION

Occupational exposure to hazardous drugs can lead to adverse effects on health care workers, including nurses, pharmacists, pharmacy technicians, and support workers.¹ In particular, exposure to hazardous drugs has been shown to lead to adverse reproductive outcomes.^{2,3} To raise awareness about this issue, the US National Institute for Occupational Safety and Health (NIOSH) published an alert on the prevention of occupational exposure to hazardous drugs in 2004.⁴ Since then, NIOSH has been updating its list of hazardous drugs every 2 years. In addition to antineoplastic drugs, other drugs that are considered hazardous include immunosuppressants, antipsychotics, antidepressants, and hormones. In the 2014 NIOSH list of hazardous drugs, the majority of drugs (97/184 [53%]) are antineoplastic drugs.⁵

Following the 2004 NIOSH alert,⁴ many organizations reviewed their guidelines for the safe handling of hazardous drugs. For instance, the American Society of Health-System Pharmacists published new guidelines in 2006.⁶ The United States Pharmacopeial Convention is also developing a new guideline (USP General Chapter <800>) that will contain specific requirements for compounding both antineoplastic and non-antineoplastic hazardous drugs.⁷ In Quebec, the Association paritaire pour la santé et la sécurité du travail du secteur des affaires sociales (a “joint, sector-based association dedicated to promoting occupational health and safety prevention and supporting health and social service sector workers and institutions”) published a safe handling guide in 2008.⁸ In addition, the Ordre des pharmaciens du Québec recently published a new guideline for compounding sterile hazardous drugs.⁹ This guideline recommends that chemical contamination on work surfaces be assessed twice per year.

Since 2008, the authors’ group has performed 3 multicentre studies of environmental contamination in Quebec hospitals.¹⁰⁻¹² For a more recent investigation, reported here, the scope of study was expanded to include Canadian centres outside Quebec. The aim of this study was to monitor environmental contamination with cyclophosphamide, ifosfamide, and methotrexate in oncology pharmacy and patient care areas in Canadian hospitals. The secondary objective was to describe temporal trends.

METHODS

Participating Hospitals

Directors of pharmacy departments in Canadian hospitals with at least 50 acute care beds were contacted by e-mail. Any hospital that handled one or more of the 3 target antineoplastic drugs was eligible to participate in the study. Pharmacy directors in Quebec hospitals ($n = 58$) were contacted on December 20, 2013, and a reminder was sent on January 10, 2014. Pharmacy directors in hospitals in other provinces ($n = 137$) were contacted

on January 10, 2014 (there was no systematic follow-up for potential respondents outside Quebec).

Each participating hospital was expected to apply local policies and procedures for compounding, administration, surface cleaning, waste management, and other aspects of drug handling. All participating hospitals were equipped with laminar air flow cabinets in the pharmacy, for sterile compounding. Each hospital assumed the cost of analysis for its own samples. Participants were asked whether they were using a closed-system transfer device (CSTD) for preparing antineoplastic drugs in the pharmacy (i.e., “a drug transfer device that mechanically prohibits the transfer of environmental contaminants into the system and the escape of hazardous drug or vapor concentrations outside the system”⁴). Participants were also asked whether they removed the outer packaging of antineoplastic drugs in the pharmacy after receipt and whether they cleaned the vials of antineoplastic drugs in the pharmacy after receipt. After the study was completed, each participating centre received a report comparing its results with global results from all participating centres. The authors’ research group previously set the overall 75th percentile of surface concentration as a goal for every participating centre; after the conclusion of the study, centres were encouraged to target sampling sites with values above this goal for corrective measures.

Sampling Technique

The sampling technique was described previously.¹⁰⁻¹² Twelve standardized measurement sites, 6 in pharmacy areas and 6 in patient care areas, were selected for sampling (Table 1). These sites were identical with those targeted in the 2008–2010, 2012, and 2013 studies.¹⁰⁻¹² Each participating hospital was given a description and photographs of the standardized sampling sites. The photographs provided were taken at the investigators’ hospital. Each participating hospital was also asked to provide a picture of the actual sites used for sampling. Samples were collected by one research assistant (A.J.) from the research team (for nearby hospitals) or by a trained employee from the participating hospital (for remote hospitals). A video of the sampling technique was provided for training of employees. For each sample, a standardized surface of about 600 cm² (20 cm × 30 cm) was sampled with one 6 cm × 8 cm Wypall X60 wipe (Kimberly Clark Professional, Newton Square, Pennsylvania). The wipe was moistened with 1 mL of sampling solution (10% methanol and 90% 5 mmol/L ammonium acetate). Sites were sampled at the end of a workday or in the morning, before surfaces were washed. The sampling technique, an adaptation of the technique described by Larson and others,¹³ was developed by the Institut national de santé publique du Québec.

Analytical Procedure

Sampling wipes were stored at a temperature between 2°C and 8°C in 50-mL polypropylene tubes. Before analysis, 10 mL

Table 1. Description of 12 Standardized Sites to be Sampled at Each Centre

Sampling Site	Description
Pharmacy areas	
Shipment reception counter	Counter used for receiving shipments and for unpacking antineoplastic drugs
Storage shelf or bin	Shelf or bin used for storage of antineoplastic drugs
Front grille inside hood	Grille located in the front of the main hood (biological safety cabinet) used for compounding antineoplastic drugs
Floor in front of hood	Floor in front of the main hood used for compounding antineoplastic drugs
Service hatch or counter for postpreparation validation	Service hatch used to transfer drugs from the compounding room to the postpreparation validation area or counter used by pharmacy personnel for postpreparation validation
Tray used for drug delivery	Tray or container used to deliver antineoplastic drugs to patient care areas after their preparation
Patient care areas	
Storage shelf or bin	Shelf or bin used for storage of antineoplastic drugs
Counter used for priming and validation	Counter used for priming tubing for antineoplastic drugs or for the nurse's final validation of compounded syringes before administration of drugs to patients; if no priming counter, a counter where the drugs are stored before administration.
Armrest	Armrest (on a chair or elsewhere) where a patient would put his or her arm during administration of an antineoplastic drug from a peripheral line
Counter in patient's room	Counter (or table) in a patient's room where drugs and related devices are placed during drug administration, in a room where at least one dose of cyclophosphamide, ifosfamide, or methotrexate was given in the 12-h period before sampling
Counter in outpatient clinic	Counter (or table) in an outpatient clinic where drugs and related devices are placed during drug administration, in a location where at least one dose of cyclophosphamide, ifosfamide, or methotrexate was given in the 12-h period before sampling
Exterior surface of antineoplastic drug container	Exterior surface of syringe or bag containing compounded drug

of extracting solution and internal standards were added to each tube. Each tube was mechanically stirred for 10 min, and an aliquot of the solution was removed for analysis. For each sample, 3 antineoplastic drugs (cyclophosphamide, ifosfamide, methotrexate) were quantified by ultra-performance liquid chromatography tandem mass spectrometry (Acquity UPLC chromatographic system coupled with Xevo TQ-S tandem mass spectrometer; Waters, Milford, Massachusetts). Chromatography was carried out on a C18 Acquity UPLC BEH column (2.1 × 50 mm, 1.7 µm; Waters, Milford, Massachusetts) with a gradient from 10/90 to 60/40 of methanol / 5 mmol/L ammonium acetate over 2 min. Overall, mean recovery from surfaces was 79%, and the intra-assay coefficient of variation was 22%. Recovery from surfaces ranged from 67% to 89% for stainless steel (coefficient of variation 5%–8%), from 79% to 102% for melamine (coefficient of variation 1%–3%), from 89% to 92% for plastic (coefficient of variation 4%–6%), and from 37% to 82% for linoleum (coefficient of variation 7%–22%).

Results expressed in nanograms per millilitre (ng/mL) were converted to nanograms per square centimetre (ng/cm²). These values were then multiplied by 11 (the dilution factor) and divided by 600 cm² (the surface area sampled) to obtain the final results, which were expressed in picograms per square centimetre (pg/cm²). The limit of detection was 0.36 pg/cm² (19.8 pg/mL) for cyclophosphamide, 0.95 pg/cm² (52 pg/mL) for ifosfamide, and 0.97 pg/cm² (53 pg/mL) for methotrexate. The limit of quantification was 1.21 pg/cm² (65.9 pg/mL) for cyclophosphamide, 3.17 pg/cm² (173 pg/mL) for ifosfamide, and 3.25 pg/cm² (177 pg/mL) for methotrexate. The limit of detection was used as the reporting limit.

Data Analysis

The proportion of positive samples was calculated. A sample was considered positive for a particular drug if the value was above the limit of detection. Descriptive statistical analyses (minimum, median, 75th percentile, 90th percentile, maximum) were carried out. For calculations, concentrations that fell between the limit of detection and the limit of quantification were assigned a value corresponding to the limit of quantification divided by 2,¹⁴ and concentrations that fell below the limit of detection were assigned a value corresponding to the limit of detection divided by 2.¹⁵

Results from the hospitals that participated in the 3 earlier studies¹⁰⁻¹² and the current study were used for comparisons over time.

Subanalyses were performed according to hospitals' working practices, i.e., use of a CSTD, removal of packaging, and cleaning of vials after initial receipt. The effect of CSTD use was evaluated for pharmacy sampling sites corresponding to steps performed during and after compounding (i.e., hood, floor, service hatch, delivery tray). The effect of packaging removal and cleaning of vials was evaluated for pharmacy sampling sites corresponding to steps performed after receipt of drugs (i.e., storage shelf, hood, floor, service hatch, delivery tray). Results were compared with a Kolmogorov–Smirnov test for 2 unpaired samples. A *p* value less than 0.05 was considered significant. For these subanalyses, only the results for cyclophosphamide contamination were used, as they were deemed representative of the current situation; there was too little surface contamination with ifosfamide and methotrexate to allow similar subanalyses.

RESULTS

Participating Hospitals

A total of 51 Canadian hospitals participated in this study: 34 (59%) of the 58 Quebec hospitals and 17 (12%) of the 137 centres from other Canadian provinces. The respondents from provinces other than Quebec were from Manitoba, New Brunswick, Nova Scotia, Ontario, and Prince Edward Island.

Two-thirds of the participating centres (34/51 [67%]) were teaching hospitals. Nearly all participating centres (49/51 [96%]) provided information about their working practices. Of these, 12 (24%) used a CSTD: ChemoClave System (ICU Medical Inc, San Clemente, California) ($n = 6$), PhaSeal (Becton, Dickinson and Company, Franklin Lakes, New Jersey) ($n = 3$), Chemo Dispensing Pin (B Braun Medical Inc, Bethlehem, Pennsylvania) ($n = 1$), unspecified ($n = 2$). Greater proportions of these hospitals removed the outer packaging after receipt (29/49 [59%]) and cleaned the vials after receipt (28/49 [57%]). Among the 49 respondents providing information about working practices, 7 (14%) used a CSTD, removed outer packaging, and cleaned vials after receipt.

Environmental Contamination with Antineoplastic Drugs

A total of 584 samples were collected between February and September 2014 (303 from pharmacy areas and 281 from patient

care areas). The median number of sites per hospital with at least one positive sample for any drug was 7 (range 1–11). All participating hospitals had at least 1 positive sample for at least 1 of the 3 antineoplastic drugs evaluated (cyclophosphamide, ifosfamide, or methotrexate).

Overall, 50% (294/584) of the samples were positive for cyclophosphamide, 21% (125/584) were positive for ifosfamide, and 9% (54/584) were positive for methotrexate (Table 2). For 6 sampling sites—the storage shelf or bin in the pharmacy, the front grille inside the hood, the floor in front of the hood, the armrest, a counter in the patient’s room, and a counter used for priming—at least half of the samples were positive for cyclophosphamide.

The overall 75th percentiles for drug concentration were 10.8 pg/cm² for cyclophosphamide, 1.59 pg/cm² for ifosfamide, and below the limit of detection for methotrexate (Table 3).

Comparison of Hospitals in Quebec and the Rest of Canada

Because participants in this study included centres located outside Quebec, the level of surface contamination was compared according to geographic location. Surface contamination with cyclophosphamide was similar in pharmacy areas in hospitals inside and outside Quebec (Figure 1). However, contamination with cyclophosphamide was higher in patient care areas from Quebec hospitals, which resulted in higher overall contamination

Table 2. Proportion of Samples Testing Positive* for Antineoplastic Drugs in Pharmacy and Patient Care Areas in 51 Canadian Hospitals (Sampling in 2014)

Sampling Site	Drug; No. (%) of Samples with Positive Result		
	Cyclophosphamide	Ifosfamide	Methotrexate
Pharmacy areas			
Shipment reception counter ($n = 50$)	9 (18)	2 (4)	4 (8)
Storage shelf or bin ($n = 50$)	25 (50)	19 (38)	10 (20)
Front grille inside hood ($n = 51$)	42 (82)	18 (35)	16 (31)
Floor in front of hood ($n = 51$)	38 (75)	23 (45)	9 (18)
Service hatch or counter for postpreparation validation ($n = 51$)	17 (33)	10 (20)	1 (2)
Trays used for drug delivery ($n = 50$)	13 (26)	10 (20)	4 (8)
Subtotal ($n = 303$)	144 (48)	82 (27)	44 (15)
Patient care areas			
Storage shelf or bin ($n = 50$)	17 (34)	6 (12)	2 (4)
Counter used for priming or validation ($n = 50$)	28 (56)	4 (8)	3 (6)
Armrest ($n = 47$)	43 (91)	15 (32)	0 (0)
Counter in patient’s room ($n = 44$)	27 (61)	11 (25)	2 (5)
Outpatient clinic counter ($n = 44$)	19 (43)	2 (5)	1 (2)
Exterior surface of antineoplastic drug container ($n = 46$)	16 (35)	5 (11)	2 (4)
Subtotal ($n = 281$)	150 (53)	43 (15)	10 (4)
Overall total ($n = 584$)	294 (50)	125 (21)	54 (9)

*A sample was considered positive if it was above the limit of detection for the particular drug. The limits of detection were 0.36 pg/cm² (19.8 pg/mL) for cyclophosphamide, 0.95 pg/cm² (52 pg/mL) for ifosfamide, and 0.97 pg/cm² (53 pg/mL) for methotrexate.

(Figure 1). The most contaminated sites were the same for all Canadian centres.

Trends from 2008–2010 to 2014

Three similar studies were conducted in Quebec in 2008–2010, 2012, and 2013, with 25, 33, and 36 participating hospitals, respectively.¹⁰⁻¹² Nineteen hospitals participated in all 4 studies. There was no difference in surface contamination between the 19 centres that participated in all 4 studies and the 51 centres that participated in the 2014 study (Figure 2). The proportion of positive samples remained constant over the years (Figure 2A),

but the 75th percentile of cyclophosphamide surface concentration declined and reached a steady state in 2012 (Figure 2B).

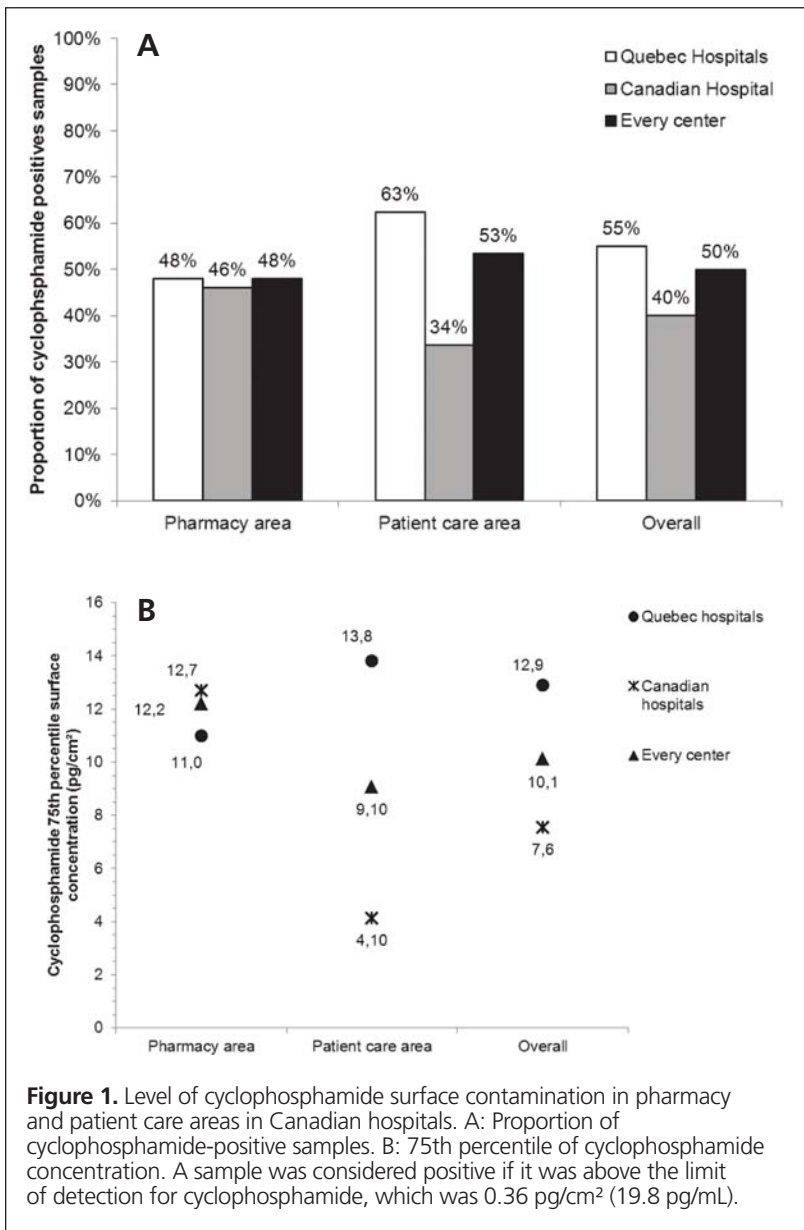
It is relevant to mention that the limits of detection declined over the years, which may have limited comparisons among the 4 studies. To test this possibility, the proportion of positive samples was recalculated using the value for limit of detection that was in effect in the 2008–2010 study, which was higher than the current limit of detection. With this change, the proportion of positive samples still remained constant over the years (data not shown). Thus, we are confident that the trend as reported here is accurate and was not caused by a change in the limits of detection.

Table 3. Surface Contamination with Antineoplastic Drugs in Pharmacy and Patient Care Areas in 51 Canadian Hospitals (Sampling in 2014)

Sample Site	Drug: Concentration (pg/cm ²)*								
	Cyclophosphamide			Ifosfamide			Methotrexate		
	50th perc.	75th perc.	Max	50th perc.	75th perc.	Max	50th perc.	75th perc.	Max
Pharmacy areas									
Shipment reception counter (n = 50)	< LOD	< LOD	86.9	< LOD	< LOD	67.9	< LOD	< LOD	22.6
Storage shelf (n = 50)	0.600	3.91	298	< LOD	3.69	202	< LOD	1.46	451
Front grille inside hood (n = 51)	21.5	153.0	3 200	< LOD	8.32	85 100	< LOD	3.36	1 080
Floor in front of hood (n = 51)	12.7	67.6	555	< LOD	9.42	455	< LOD	< LOD	13.0
Service hatch or counter for post-preparation validation (n = 51)	< LOD	3.21	3 850	< LOD	< LOD	1 050	< LOD	< LOD	4.46
Trays used for drug delivery (n = 50)	< LOD	0.61	53.2	< LOD	< LOD	15 400	< LOD	< LOD	28.1
Subtotal (n = 303)	0.360	12.2	3 850	< LOD	1.65	85 100	< LOD	< LOD	1 080
Patient care areas									
Storage shelf (n = 50)	< LOD	1.28	11 430	< LOD	< LOD	47.8	< LOD	< LOD	52.1
Counter used for priming or validation (n = 50)	0.6	4.77	82.3	< LOD	< LOD	23.6	< LOD	< LOD	13.3
Armrest (n = 47)	45.3	159	903	< LOD	5.75	1 050	< LOD	< LOD	< LOD
Patient room counter (n = 44)	2.68	6.88	38.5	< LOD	< LOD	512	< LOD	< LOD	1.62
Outpatient clinic counter (n = 44)	< LOD	3.27	27.7	< LOD	< LOD	35.7	< LOD	< LOD	10.0
Exterior surface of antineoplastic drug container (n = 46)	1.51	8.04	8 290	2.15	3.97	233	2.19	4.05	249
Subtotal (n = 281)	1.51	9.06	11 400	< LOD	1.43	1 050	< LOD	< LOD	249
Overall total (n = 584)	0.72	10.8	11 400	< LOD	1.59	85 100	< LOD	< LOD	1 080

LOD = limit of detection, Max = maximum, perc. = percentile.

*The LOD was 0.36 pg/cm² (19.8 pg/mL) for cyclophosphamide, 0.95 pg/cm² (52 pg/mL) for ifosfamide, and 0.97 pg/cm² (53 pg/mL) for methotrexate.



Over the years, the most frequent cyclophosphamide-positive sampling sites in the pharmacy were the front grille of the hood and the floor in front of the hood (Figure 3A). In patient care areas, the most frequent cyclophosphamide-positive sampling site was the armrest (Figure 3B).

Effects of Working Practices

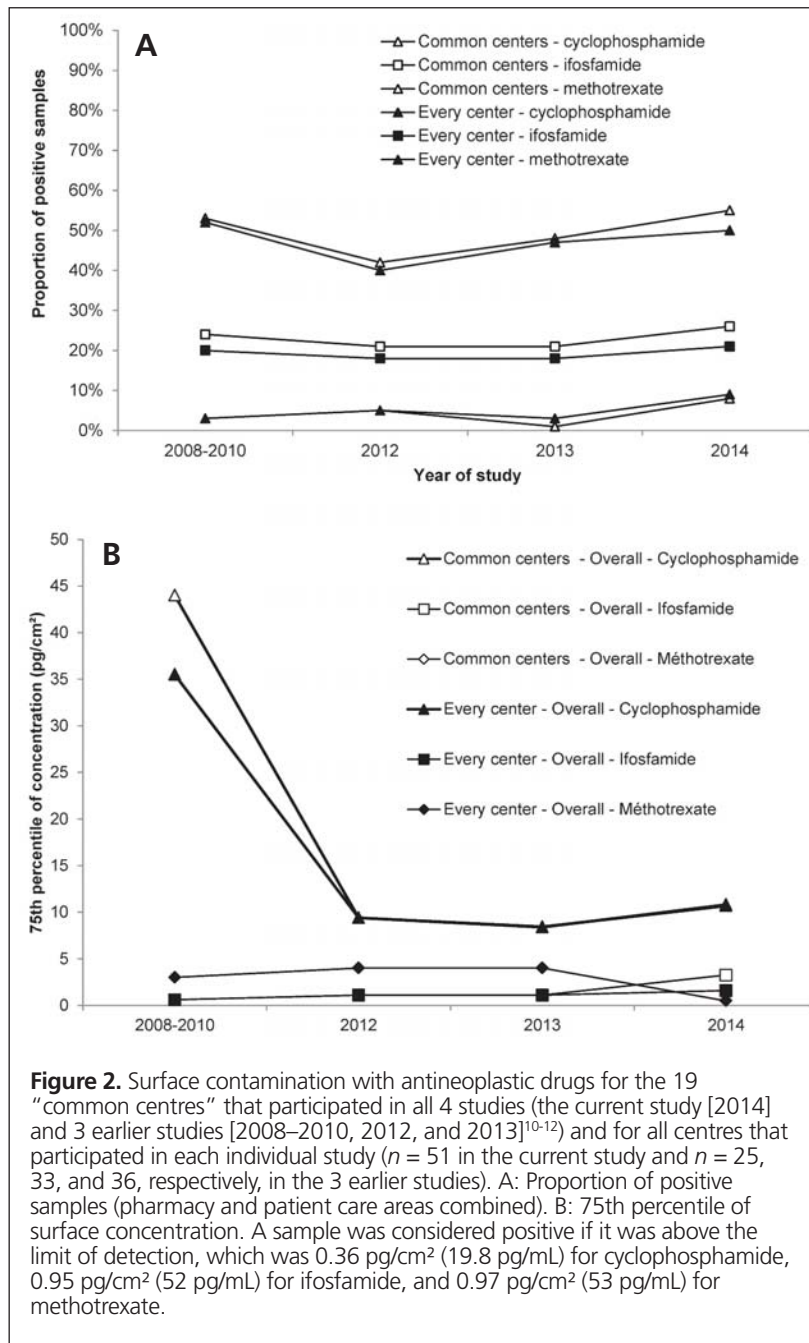
The potential link between certain working practices and surface concentration with cyclophosphamide in the pharmacy was investigated. Overall, the 75th percentile for cyclophosphamide concentration was lower for centres that used a CSTD, removed the outer packaging of vials after receipt, and/or cleaned

vials after receipt, but this difference was not significant (Tables 4–6).

DISCUSSION

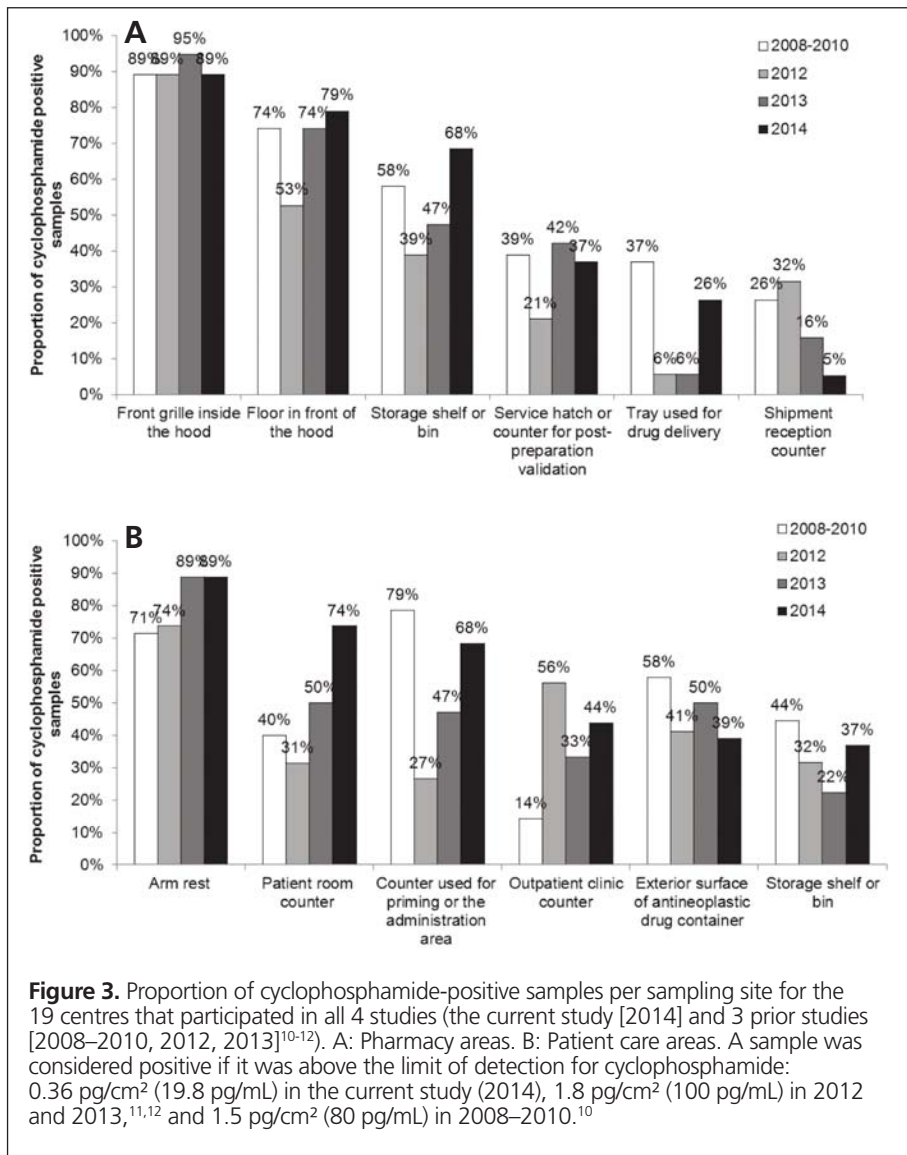
Environmental Monitoring

Overall, among samples obtained from 51 Canadian hospitals participating in this study, 50% were positive for cyclophosphamide, 21% were positive for ifosfamide, and 9% were positive for methotrexate. The 75th percentile values for surface concentration were 10.8 pg/cm² for cyclophosphamide, 1.59 pg/cm² for ifosfamide, and below the limit of detection for methotrexate.



Relative to 3 other multicentre studies conducted within Quebec in 2008–2010, 2012, and 2013, respectively,¹⁰⁻¹² the proportion of positive samples in the current study remained constant. Nonetheless, the surface concentration of antineoplastic drugs measured has been decreasing over the years and seems to have reached a plateau in 2012.

This reduction in surface contamination can probably be explained by an increase in awareness of the importance of safe-handling practices, following the adoption of improved local procedures^{8,9} and the publication of international guidelines.^{6,7} It will be interesting to find out, over the coming years, whether the new guideline for compounding hazardous drugs of the



Ordre des pharmaciens du Québec⁹ will have an effect on surface contamination.

Sampling Sites

Over the 4 studies, the most frequent cyclophosphamide-positive sample sites were the front grille of the hood, the floor in front of the hood, and the armrest of the chair used during administration of antineoplastic drugs. Similar sites were found to be highly contaminated in another recent large study involving 30 US hospitals. In that study, Sessink and others¹⁶ found that 97% of samples from the front grille of the hood and 82% of those from the floor in front of the hood were positive for cyclophosphamide.

These frequently contaminated sites correspond to sites where large quantities of antineoplastic drugs are manipulated.

In a Canadian study, Hon and others¹⁷ identified the steps of drug preparation and drug administration as critical aspects of the medication-use system. In addition to sites where large quantities of antineoplastic drugs are manipulated, these authors found contamination on everyday objects such as pencils and door handles.¹⁷ Indeed, skin absorption and inhalation are frequent sources of contamination, but hand-to-mouth contact also leads to occupational exposure.³ In another Canadian study, Hon and others¹⁸ evaluated the contamination of hands of health care workers. They found that workers in the patient care unit who were not directly involved in drug administration, such as dietitians, ward aides, oncologists, and volunteers, had the highest rate of hand contamination, with 28.6% of this group testing positive for contamination.¹⁸

In addition to emphasizing the importance of adequate working practices and personal protective equipment, the

Table 4. Effect of Using Closed-System Transfer Devices (CSTDs) on Contamination of Selected Sampling Sites in the Pharmacy

Sampling Site	75th Percentile of Cyclophosphamide Concentration (pg/cm ²)		p Value*
	CSTD in Use (n = 12)	CSTD Not in Use (n = 37)	
Front grille inside hood	112	193	0.40
Floor in front of hood	48.6	82.6	0.43
Service hatch or counter for postpreparation validation	0.182	3.35	0.41
Tray used for drug delivery	0.303	2.44	0.46

*Kolmogorov–Smirnov test for 2 unpaired samples.

Table 5. Effect of Removing Outer Vial Packaging on Contamination of Selected Sampling Sites in the Pharmacy

Sampling Site	75th Percentile of Cyclophosphamide Concentration (pg/cm ²)		p Value*
	Removal of Outer Packaging (n = 29)	No Removal of Outer Packaging (n = 20)	
Storage shelf or bin	3.90	3.05	0.41
Front grille inside hood	137	173	0.50
Floor in front of hood	61.9	94.8	0.48
Service hatch or counter for post-preparation validation	2.89	4.05	0.50
Tray used for drug delivery	0.363	4.32	0.18

*Kolmogorov–Smirnov test for 2 unpaired samples.

Table 6. Effect of Cleaning Vials on Contamination of Selected Sampling Sites in the Pharmacy

Sampling Site	75th Percentile of Cyclophosphamide Concentration (pg/cm ²)		p Value*
	Cleaning of Vial Exterior (n = 28)	No Cleaning of Vial Exterior (n = 21)	
Storage shelf or bin	3.40	4.41	0.48
Front grille inside hood	151	169	0.32
Floor in front of hood	64.7	132	0.32
Service hatch or counter for post-preparation validation	2.09	3.35	0.50
Tray used for drug delivery	0.272	4.28	0.19

*Kolmogorov–Smirnov test for 2 unpaired samples.

presence of contamination highlights the usefulness of suitable cleaning methods. The use of alcohol to clean a surface can often spread the contamination, rather than eliminating it.¹⁹ The use of detergent and water is better for cleaning a surface, but to totally eliminate all traces of contaminants, decontamination should be done with a combination of sodium hypochlorite and sodium thiosulfate.²⁰

Pharmacy and Patient Care Areas

Overall, surface contamination in the current study was similar in pharmacy and patient care areas. However, contamination of patient care areas with cyclophosphamide was higher for Quebec hospitals than for hospitals from other provinces. Although the data collected for this study were insufficient to

explain this difference, it would be interesting to determine whether different practices for activities such as cleaning, drug preparation, and drug administration are used in patient care units in Quebec and the other provinces. The difference in response rates between Quebec and the rest of Canada limits this comparison, and participating hospitals may not be representative of all Canadian hospitals.

Effects of Working Practices

Subanalyses were performed to evaluate the effect of 3 working practices: use of a CSTD, removal of packaging, and cleaning of vials after initial receipt. No statistically significant differences were found between hospitals that did and did not follow these practices. In the 2013 study,¹² the concentration of contaminants

was lower on the front grille of the hood at centres that used CSTDs, removed the outer packaging, and cleaned vials after receipt. Even though numerous studies have shown that CSTDs can reduce contamination, their use does not completely eliminate contamination. For instance, Sessink and others¹⁶ found a median of 1.69 ng/cm² (or 1690 pg/cm²) of cyclophosphamide on the front grille of the hood before CSTDs were in use and 0.39 ng/cm² (or 390 pg/cm²) after CSTDs were implemented. In the current study, the median concentration of cyclophosphamide on the front grille of the hood was 43 pg/cm² at hospitals not using a CSTD ($n = 12$) and 15 pg/cm² at hospitals that did use such devices ($n = 37$). As such, hospitals in this study that did not use CSTDs had a 26-fold lower concentration of cyclophosphamide than hospitals in other studies that did use CSTDs (15 pg/cm² versus 390 pg/cm²). The very low surface contamination observed in the current multicentre study, despite the fact that few participating hospitals (12/49) reported the use of CSTDs, indicates that many strategies can contribute to a low level of surface contamination.

It is recommended that vials be unpacked and cleaned after initial receipt.⁸ Although we did not find a significant difference between hospitals that did and did not follow this practice, this low-cost solution can probably help to reduce contamination, given that the exterior surface of vials is often contaminated during the manufacturing process.²¹

Implications for Practice

As long as no health-based safe limit of exposure is known, workers' exposure to antineoplastic and other hazardous drugs should be kept as low as reasonably achievable. We suggest the use of local (country- or region-specific) and attainable goals and recommend that centres strive to attain a level of contamination lower than these targets. For each drug, the value of the most recent global 75th percentile (derived from all centres that participated in the current study) should be used as a manageable target for Canadian hospitals. It is hoped that these target values will continue to be reduced over the years to come.

How can hospitals reduce their level of contamination? Environmental sampling can help to identify problem areas, which can then be cleaned thoroughly with a combination of sodium hypochlorite and sodium thiosulfate. Such cleaning should be done regularly, and a log should be signed by the workers responsible, especially for areas where large quantities of antineoplastic agents are manipulated (e.g., the hood in the pharmacy) and areas that are touched frequently (e.g., armrests). The source of the contamination may be the exterior of vials, so cleaning vials when they are received and cleaning the container for the compounded product once prepared can also limit contamination. Materials that cannot be cleaned properly should be substituted with materials that can be cleaned, if possible.

Finally, it is important to continue raising the awareness of all workers who are potentially exposed to antineoplastic drugs,

to ensure that they use proper working practices that will limit the spread of contamination and that they adequately protect themselves with personal protective equipment.⁸

Strengths and Limitations

This study was the largest multicentre study of hazardous-drug contamination to date in Canadian hospitals and provides results for both pharmacy and patient care areas. To limit technical bias and to ensure a consistent sampling method across all centres, the investigators supplied a video demonstrating the correct sampling technique. As much as possible, sampling was performed at the end of the day, to generate values that were representative of a working day and also representative of the potential professional exposure to these drugs among health care workers. However, all sampling at each institution was performed on a single day, and different results might have been obtained on a different day. Many different analytical techniques are available, so caution should be used when comparing these results with the results of other studies. The limits of detection were comparable to those used by other investigators. The global recovery rate from surfaces was adequate; however, the recovery from linoleum (floors) was lower, so those results might be underestimations. The cost of the analysis may have prevented some hospitals from participating.

CONCLUSIONS

Relative to 3 other multicentre studies conducted in Quebec over the past several years, the proportion of positive samples remained constant. Nonetheless, the 75th percentile surface concentration of antineoplastic drugs has been decreasing over time and seems to have reached a plateau. As long as no health-based limit of exposure is known, local (country- or region-specific) and attainable goals for surface contamination with hazardous drugs should be set annually.

References

1. Medical surveillance for healthcare workers exposed to hazardous drugs. Cincinnati (OH): Department of Health and Human Services (US), Centers for Disease Control and Prevention, National Institute for Occupational Safety and Health; 2012 [cited 2013 Oct 15]. Available from: www.cdc.gov/niosh/docs/wp-solutions/2013-103/pdfs/2013-103.pdf
2. Dranitsaris G, Johnston M, Poirier S, Schueller T, Milliken D, Green E, et al. Are health care providers who work with cancer drugs at an increased risk for toxic events? A systematic review and meta-analysis of the literature. *J Oncol Pharm Pract.* 2005;11(2):69-78.
3. Connor TH, Lawson CC, Polovich M, McDiarmid MA. Reproductive health risks associated with occupational exposures to antineoplastic drugs in health care settings: a review of the evidence. *J Occup Environ Med.* 2014;56(9):901-10.
4. *Preventing occupational exposure to antineoplastic and other hazardous drugs in healthcare settings.* Publ No. 2004-165. Cincinnati (OH): Department of Health and Human Services (US), Centers for Disease Control and Prevention, National Institute for Occupational Safety and Health; 2004 [cited 2014 Oct 24]. Available from: www.cdc.gov/niosh/docs/2004-165/
5. Connor TH, MacKenzie BA, DeBord DG, Trout DB, O'Callaghan JP. *NIOSH list of antineoplastic and other hazardous drugs in healthcare settings, 2014.* Cincinnati (OH): Department of Health and Human Services (US),

- Centers for Disease Control and Prevention, National Institute for Occupational Safety and Health; 2014 [cited 2014 Oct 24]. Available from: www.cdc.gov/niosh/docs/2014-138/pdfs/2014-138.pdf
- American Society of Health-System Pharmacists. ASHP guidelines on handling hazardous drugs. *Am J Health Syst Pharm.* 2006;63(12):1172-93.
 - <800> Hazardous drugs—handling in healthcare settings [briefing]. Rockville (MD): U.S. Pharmacopeial Convention; 2014 [cited 2014 Oct 24]. Available from: www.usp.org/sites/default/files/usp_pdf/EN/m7808.pdf
 - Prevention guide—safe handling of hazardous drugs*. Montréal (QC): Association paritaire pour la santé et la sécurité du travail du secteur des affaires sociales; 2008 [cited 2014 Oct 24]. Available from: www.asstas.qc.ca/publications/publications-specialisees/guides-de-prevention/prevention-guide-safe-handling-of-hazardous-drugs.html
 - Norme 2014.02 : Préparation de produits stériles dangereux en pharmacie. Montréal (QC): Ordre des pharmaciens du Québec; 2014 [cited 2014 Mar 20]. Available from: www.opq.org/fr-CA/publications/normes-de-pratique-et-lignes-directrices/
 - Bussièrès JF, Tanguay C, Touzin K, Langlois É, Lefebvre M. Environmental contamination with hazardous drugs in Quebec hospitals. *Can J Hosp Pharm.* 2012;65(6):428-35.
 - Merger D, Tanguay C, Langlois E, Lefebvre M, Bussièrès JF. Multicenter study of environmental contamination with antineoplastic drugs in 33 Canadian hospitals. *Int Arch Occup Environ Health.* 2014;87(3):307-13.
 - Berruyer M, Tanguay C, Caron NJ, Lefebvre M, Bussièrès JF. Multicenter study of environmental contamination with antineoplastic drugs in 36 Canadian hospitals: a 2013 follow-up study. *J Occup Environ Hyg.* 2015;12(2):87-94.
 - Larson RR, Khazaeli MB, Dillon HK. Monitoring method for surface contamination caused by selected antineoplastic agents. *Am J Health Syst Pharm.* 2002;59(3):270-7.
 - Article 5: calculation of mean values. In: Commission directive 2009/90/CE of 31 July 2009 laying down, pursuant to Directive 2000/60/EC of the European Parliament and of the Council, technical specifications for chemical analysis and monitoring of water status. *Official Journal of the European Union*. Brussels (Belgium): European Union; 2009 [cited 2014 Oct 24]. Available from: <http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2009:201:0036:0038:EN:PDF>
 - Hornung RW, Reed LD. Estimation of average concentration in the presence of nondetectable values. *Appl Occup Environ Hyg.* 1990;5(1):46-51.
 - Sessink PJ, Trahan J, Coyne W. Reduction in surface contamination with cyclophosphamide in 30 US hospital pharmacies following implementation of a closed-system drug transfer device. *Hosp Pharm.* 2013;48(3):204-12.
 - Hon CY, Teschke K, Chu W, Demers P, Venners S. Antineoplastic drug contamination of surfaces throughout the hospital medication system in Canadian hospitals. *J Occup Environ Hyg.* 2013;10(7):374-83.
 - Hon CY, Teschke K, Demers PA, Venners S. Antineoplastic drug contamination on the hands of employees working throughout the hospital medication system. *Ann Occup Hyg.* 2014;58(6):761-70.
 - Chu WC, Hon CY, Danyluk Q, Chua PP, Astrakianakis G. Pilot assessment of the antineoplastic drug contamination levels in British Columbian hospitals pre- and post-cleaning. *J Oncol Pharm Pract.* 2012;18(1):46-51.
 - Touzin K, Bussièrès JF, Langlois E, Lefebvre M, Métra A. Pilot study comparing the efficacy of two cleaning techniques in reducing environmental contamination with cyclophosphamide. *Ann Occup Hyg.* 2010;54(3):351-9.
 - Schierl R, Herwig A, Pfaller A, Groebmair S, Fischer E. Surface contamination of antineoplastic drug vials: comparison of unprotected and protected vials. *Am J Health Syst Pharm.* 2010;67(6):428-9.

Alexia Janes is a Research Assistant in the Pharmacy Practice Research Unit and the Pharmacy Department, Centre hospitalier universitaire Sainte-Justine, Montréal, Quebec. She is also a DPharm candidate with the Faculté des sciences pharmaceutiques et biologiques de Lille, Lille, France.

Cynthia Tanguay, BSc, MSc, is a Research Assistant in the Pharmacy Practice Research Unit and the Pharmacy Department, Centre hospitalier universitaire Sainte-Justine, Montréal, Quebec.

Nicolas J Caron, PhD, is a Biochemist with the Centre de toxicologie du Québec, Institut national de santé publique du Québec, Québec, Quebec.

Jean-François Bussièrès, BPharm, MSc, MBA, FCSHP, is Director of the Pharmacy Practice Research Unit and the Pharmacy Department, Centre hospitalier universitaire Sainte-Justine, and a Clinical Professor, Faculty of Pharmacy, Université de Montréal, Montréal, Quebec.

Competing interests: None declared.

Address correspondence to:

Jean-François Bussièrès
Pharmacy Department
Centre hospitalier universitaire Sainte-Justine
3175, chemin de la Côte Sainte-Catherine
Montréal QC H3T 1C5

e-mail: jf.bussieres@ssss.gouv.qc.ca

Acknowledgements: The authors would like to thank the 51 health care centres that participated in the 2014 study.

Funding: No funding was received for this study.